

A NEW SYNTHETIC APPROACH TO AMINOGLYCOSIDE ANTIBIOTICS  
BY USE OF OXIDATIVE DECARBOXYLATION AND REDUCTIVE  
DEACETOXYLATION AS KEY-REACTIONS

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Summary: By use of a decarboxylation reaction with lead tetraacetate and a de-acetoxylation reaction with sodium borohydride as key-reactions, paromamine (12) and tri-N-benzyloxycarbonylparomamine (12a) were synthesized from D-glucosamine.

Recently, by means of the oxidative decarboxylation reaction effected by lead tetraacetate or anodic oxidation, we developed two versatile conversion methods from carbohydrates leading to cyclitols.<sup>1,2)</sup> Taking advantage of these chemical modification methods, several successful conversions from N-acetyl-D-glucosamine to streptomine hexaacetate<sup>3,4)</sup> and from glucuronide-saponins to aminocyclitol-oligoglycosides<sup>2)</sup> were accomplished. Furthermore, in the studies on the chemical behavior of nitrocyclitol derivatives, we found that NaBH<sub>4</sub> effected one-step reductive elimination of the acetoxyl function(s) located at  $\beta$  to the nitro group. By use of this reaction, deoxyaminocyclitols (e.g. 2-deoxystreptomine) were readily synthesized from N-acetyl-D-glucosamine and formal syntheses of (-)-shikimic acid and (-)-quinic acid from D-mannose were attained.<sup>5)</sup>

As a continuing study on these chemical modifications of carbohydrates, a disaccharide (3), which was synthesized from D-glucosamine, was successfully converted to paromamine (12)<sup>6)</sup> and its tri-N-benzyloxycarbonyl (Cbz) derivative (12a)<sup>7)</sup> by utilizing Pb(OAc)<sub>4</sub> decarboxylation and NaBH<sub>4</sub> deacetoxylation as the key-reactions as described below. Since 12a was already converted to kanamycin C (13),<sup>7)</sup> the present work constituted a formal synthesis of kanamycin C from D-glucosamine.<sup>8)</sup>

Glycosidation under a N<sub>2</sub> atmosphere of 1<sup>9)</sup> (1 mole) with 2<sup>9)</sup> (2 mole) in benzene-dioxane (2:1) in the presence of *sym*-collidine and AgClO<sub>4</sub> (r.t., 30 min) furnished an  $\alpha$ -disaccharide (3, 88%),<sup>10)</sup> mp 164-165°C, C<sub>40</sub>H<sub>44</sub>N<sub>4</sub>O<sub>18</sub>,<sup>11)</sup> [ $\alpha$ ]<sub>D</sub> +8.3°, IR (CHCl<sub>3</sub>, cm<sup>-1</sup>): 3435, 3333 (NH), 1744 (br, Ac, Cbz), 1615, 1592 (arom.) 1503 (amide II, NO<sub>2</sub>, arom.), 1338 (NO<sub>2</sub>), FD-MS (*m/z*): 868 (M<sup>+</sup>) and 4 (trace), mp 107-108°C, C<sub>40</sub>H<sub>44</sub>N<sub>4</sub>O<sub>18</sub>, [ $\alpha$ ]<sub>D</sub> -17.8°, IR (CHCl<sub>3</sub>): 3433, 3320, 1736 (br), 1621, 1595, 1506, 1337, FD-MS: 868 (M<sup>+</sup>).<sup>12)</sup>

The  $\alpha$ -glycosidic linkage in the major disaccharide (3) was suggested from the glycosidation conditions<sup>13)</sup> and was substantiated by  $^1\text{H}$  NMR analysis ( $\text{d}_6$ -acetone)<sup>14)</sup> of the deacetylated product (3a), mp 183-184°C,  $\text{C}_{34}\text{H}_{38}\text{N}_4\text{O}_{15}$ , IR ( $\text{CHCl}_3$ ): 3580 (br, OH), 3440, 3347, 1700, FD-MS: 742 ( $\text{M}^+$ ), which was quantitatively prepared from 3 with 0.01% NaOMe-dry MeOH (r.t., 2 hr). Signals due to two anomeric protons were observed at  $\delta$  4.72 (1H, d,  $J = 2$  Hz) and 5.24 (1H, d,  $J = 3$  Hz). The structure 3 was further supported from  $^{13}\text{C}$  NMR data for 3a and physicochemical properties of 4a, mp 135-137°C,  $\text{C}_{34}\text{H}_{38}\text{N}_4\text{O}_{15} \cdot 1/2\text{H}_2\text{O}$ , which was prepared by deacetylation of the minor product 4.

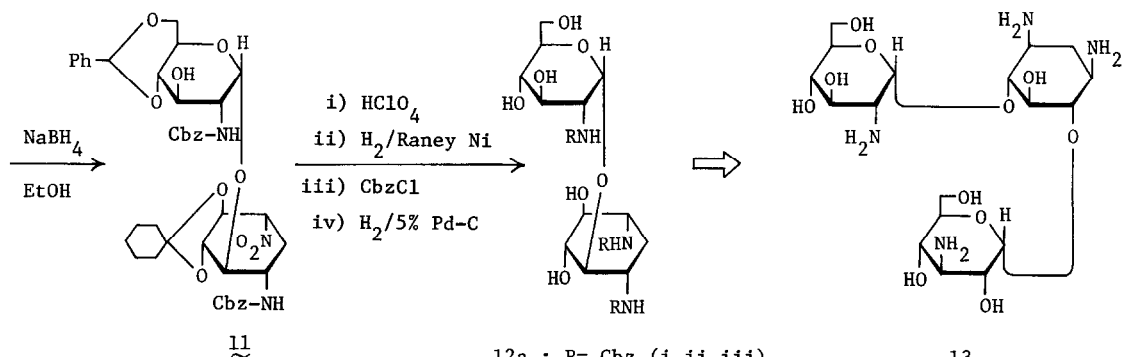
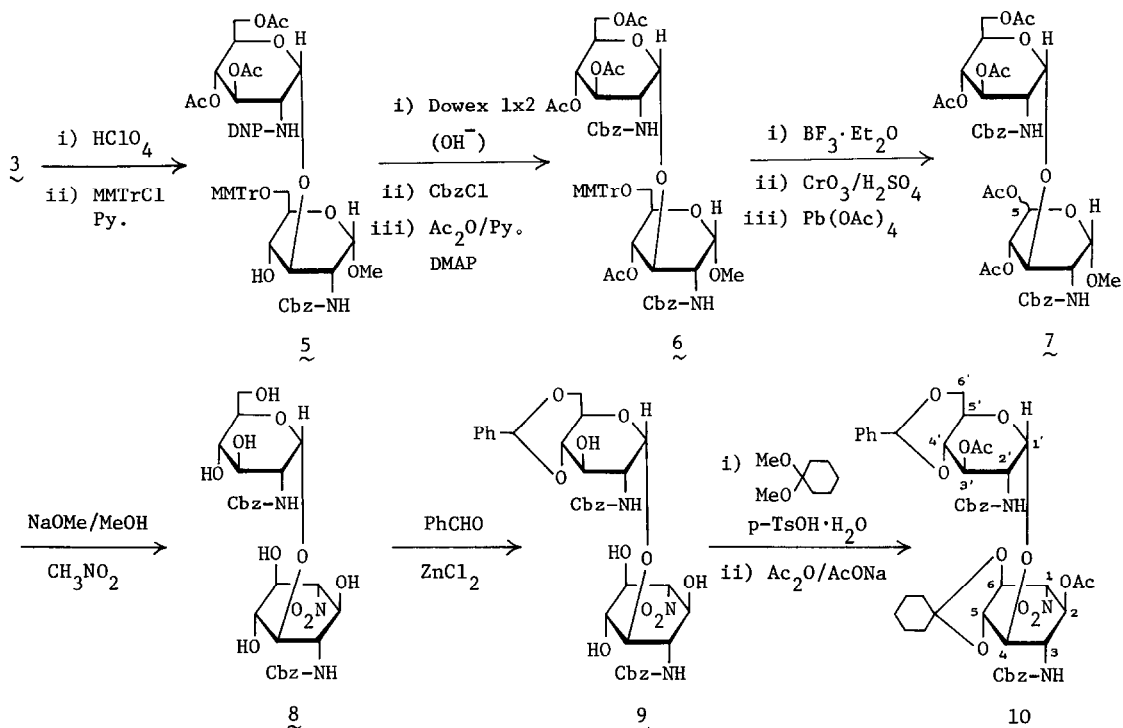
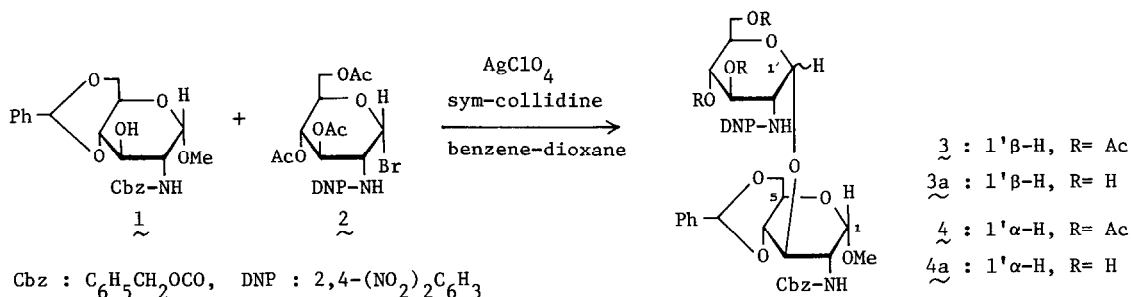
Removal of the benzyldiene group from 3 ( $\text{HClO}_4$ -acetone, r.t., 4 hr) and subsequent reaction with *p*-anisylchlorodiphenylmethane (MMTrCl)-pyridine (r.t., 2 hr) quantitatively yielded 5, mp 102-104°C,  $\text{C}_{53}\text{H}_{56}\text{N}_4\text{O}_{19} \cdot \text{H}_2\text{O}$ . The monoMMTr derivative (5) was then converted to 6 (82%), colorless oil,  $\text{C}_{57}\text{H}_{62}\text{N}_2\text{O}_{18}$ , *via* successive three reactions: i) removal of the DNP group with Dowex 1x2 ( $\text{OH}^-$ ) in acetone (r.t., 12 hr), ii) treatment with  $\text{CbzCl}$ -aq. sat.  $\text{NaHCO}_3$  in dioxane (r.t., 80 min) and iii) acetylation with  $\text{Ac}_2\text{O}$ -pyridine in the presence of 4-dimethylaminopyridine (r.t., 12 hr).

After removing the MMTr group from 6 with  $\text{BF}_3$ -etherate in dry THF-ether (1:2) (r.t., 1 hr), the alcohol ( $5\text{-CH}_2\text{OH}$ ) was oxidized with the Jones reagent (r.t., 3 hr) and the resulting uronic acid ( $5\text{-COOH}$ ) was subjected to  $\text{Pb}(\text{OAc})_4$  degradation in benzene-pyridine (50:1) (reflux, 3.5 hr)<sup>1,3)</sup> to furnish 7 (a mixture of  $5\beta$ - and  $5\alpha$ -OAc derivatives, 71% from 6), colorless oil, IR ( $\text{CHCl}_3$ ): no OH, 3439 (NH), 1745, 1232 (OAc).

Treatment of this 5-acetoxyated mixture with  $\text{CH}_3\text{NO}_2$  in 1% NaOMe-dry MeOH (r.t., 36 hr) furnished the *scyllo*-nitrocyclitol glycoside (8, 22%), very hygroscopic white powder, IR (KBr): 3369 (br), 1554, 1382 ( $\text{NO}_2$ ), FD-MS: 638 ( $\text{M}^+ + 1$ ), 660 ( $\text{M}^+ + \text{Na}$ ), together with a mixture of nitrocyclitol glycosides of other types (29%). The *scyllo* glycoside (8) was then treated with benzaldehyde and  $\text{ZnCl}_2$  under a  $\text{N}_2$  atmosphere (r.t., 2.5 hr) to furnish the monobenzyldiene derivative (9, 84%), mp 223-226°C,  $\text{C}_{35}\text{H}_{39}\text{N}_3\text{O}_{14}$ , FD-MS: 725 ( $\text{M}^+$ ). Cyclohexylidenation of 9 with 1,1-dimethoxycyclohexane and *p*- $\text{TsOH} \cdot \text{H}_2\text{O}$  in DMF (80°C, 2.5 hr) and subsequent acetylation with  $\text{Ac}_2\text{O}$ -AcONa (r.t., 30 hr) yielded 10 (52%), colorless oil,  $\text{C}_{45}\text{H}_{51}\text{N}_3\text{O}_{16}$ , IR ( $\text{CCl}_4$ ): no OH, 1742, 1727, FD-MS: 889 ( $\text{M}^+$ ).

The *scyllo* configuration of the cyclitol moiety in 10 was shown by its  $^1\text{H}$  NMR spectrum.<sup>14)</sup> Thus, in addition to proton signals due to the D-glucosamine moiety [*e.g.*  $\delta$  4.14 (1H, dd,  $J = 9, 9$  Hz,  $4'\text{-H}$ ), 5.17 (1H, d,  $J = 2$  Hz,  $1'\text{-H}$ ), and 5.40 (1H, dd,  $J = 9, 9$  Hz,  $3'\text{-H}$ )], signals assignable to the cyclitol-protons were observed at  $\delta$  4.02 (1H, dd,  $J = 10, 10$  Hz, 6-H), 4.74 (1H, dd,  $J = 10, 10$  Hz, 1-H), and 5.68 (1H, dd,  $J = 10, 10$  Hz, 2-H), which were in good accord with the *scyllo* configuration.

Reductive deacetoxylation of 10 with  $\text{NaBH}_4$  in 95% EtOH (r.t., 3.5 hr)<sup>5)</sup> furnished 11 (89%), mp 139-141°C,  $\text{C}_{41}\text{H}_{47}\text{N}_3\text{O}_{13}$ , IR ( $\text{CHCl}_3$ ): 3600, 3424, 1722,



1564, 1513, 1367, FD-MS: 789 ( $M^+$ ). The  $^1\text{H}$  NMR spectrum of 11 clearly indicated correctness of the structure:  $\delta$  1.90-2.23 (2H, m, 2- $\text{H}_2$ ) and  $\delta$  4.53 (1H, ddd,  $J=5$ , 11, 11 Hz, 1-H).

Finally, removal of the benzylidene and cyclohexylidene protecting groups from 11 with  $\text{HClO}_4$ -acetone (r.t., 10 min) and subsequent reduction (Raney Ni T-4 in  $\text{AcOH-EtOH}$ ) and benzyloxycarbonylation ( $\text{CbzCl-5\% aq. Na}_2\text{CO}_3$  in acetone) furnished tri-N-Cbz-paromamine (12a)<sup>7)</sup> (55% from 11). On the other hand, treatment of 11 with  $\text{HClO}_4$ -acetone followed by reduction of the nitro group and removal of the Cbz group by catalytic hydrogenation ( $\text{H}_2/5\% \text{ Pd-C}$ ) afforded paromamine (12) in 56% yield.

The present approach to the construction of the aminocyclitol glycoside (*e.g.* 12) is characteristic, since the conversion starts with an initial synthesis of an appropriate disaccharide and is followed by modification of one of two monosaccharide moieties into an aminocyclitol counterpart (*e.g.* the 2-deoxystreptamine moiety in 12). We are currently exploring this approach to the synthesis of other aminoglycoside antibiotics comprising more monosaccharide moieties.

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#### References and Notes

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- 11) The molecular compositions of compounds given with the chemical formulae were determined by elemental analyses.
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- 14) The  $^1\text{H}$  NMR spectra were taken on a JEOL FX-200 (200 MHz) spectrometer and the assignments were made on the basis of decoupling experiments in detail.

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